

# The Inhibition of Clostridium chauvoei (Jakari strain) Neuraminidase Activity by Methanolic Extracts of the Stem Barks of Tamarindus indicus and Combretum fragrans

N.M. USEHa,\*, A.J. NOKb, S.F. AMBALIc and K.A.N. ESIEVO

<sup>a</sup>Department of Veterinary Pathology and Microbiology, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, Nigeria; <sup>b</sup>Department of Biochemistry, Ahmadu Bello University, Zaria, Nigeria; <sup>c</sup>Department of Veterinary Physiology and Pharmacology, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, Nigeria

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The inhibition of neuraminidase from Clostridium chauvoei (jakari strain) with partially purified methanolic extracts of some plants used in Ethnopharmacological practice was evaluated. Extracts of two medicinal plants, Tamarindus indicus and Combretum fragrans at 100-1000 µg/ml, both significantly reduced the activity of the enzyme in a dose-dependent fashion (P < 0.001).

The estimated IC<sub>50</sub> values for Tamarindus indicus and Combretum fragrans were 100 and 150 µg/ml respectively. Initial velocity studies conducted, using fetuin as substrate revealed a non-competitive inhibition with the  $V_{max}$  significantly altered from 500  $\mu$ mole min<sup>-1</sup> mg<sup>-1</sup> to 240  $\mu$ mole min<sup>-1</sup> mg<sup>-1</sup> and 340  $\mu$ mole min<sup>-1</sup> mg<sup>-1</sup> in the presence of Tamarindus indicus and Combretum fragrans respectively. The K<sub>M</sub> remained unchanged at 0.42 mM. The computed Index of physiological efficiency was reduced from 1.19 min<sup>-1</sup> to 0.57 min<sup>-1</sup> and 0.75 min<sup>-</sup> with Tamarindus indicus and Combretum fragrans as inhibitors respectively.

Keywords: Tamarindus indicus and Combretum fragrans; Neuraminidase inhibition

## INTRODUCTION

Blackleg is a disease of cattle, sheep and other ruminants caused by Clostridium chauvoei.1 In Nigeria, the disease was first reported in 1929 and has remained a major problem to cattle in the country.<sup>2</sup> Neuraminidases (sialidases, EC 3.2.1.18) are involved in the pathogenesis of some infectious diseases, whose aetiologic agents produce the enzyme.3,4 The enzyme is of great importance in medicine and the pharmaceutical industry for

the analysis of oligosaccharides and development of neuraminidase inhibitors.<sup>5</sup> Inhibitors of neuraminidase are central in the clinical management of some infectious diseases, such as human influenza virus infections.6 Clostridium chauvoei (jakari strain) which causes blackleg infection in indigenous Nigerian cattle is known to produce neuraminidase and the enzyme is involved in the pathogenesis of blackleg infection by spreading the disease in host tissues.7 The nomadic Fulani pastoralists of rural Nigeria, who own about 70–80% of livestock in the country, prefer the use of herbal remedies to manage livestock diseases.8 In the present study, the effect of two medicinal plants of Nigeria (Tamarindus indicus and Combretum fragrans) used by nomads of rural Nigeria to manage blackleg9 was tested on Clostridium chauvoei (jakari strain) neuraminidase in vitro. In this report we show for the first time, the inhibition of Clostridium chauvoei (jakari strain) neuraminidase by Tamarindus indicus and Combretum fragrans.

## MATERIALS AND METHODS

# Plant Collection, Extraction and Preparation

The stem barks of both Tamarindus indicus and Combretum fragrans used in this study were collected from Samaru village, Zaria, Nigeria in September and identified at the herbarium of the Department of Biological Sciences, Ahmadu Bello University, Zaria, Nigeria. They were air dried and subsequently ground to powder. The powder materials of each of



<sup>\*</sup>Corresponding author. Tel.: +234-069-551358. Fax: +234-0803-7032523. E-mail: nickuseh@yahoo.com

340 N.M. USEH et al.

these plants were weighed and then individually mixed with methanol in a ratio of 1:5 w/v in an Erlenmeyer flask for 24 h. The mixture was filtered and the filtrate concentrated in vacuo at 50°C in a rotatory evaporator coupled to a thermocirculator. The resultant extract of each of these plants was weighed and reconstituted with distilled water to obtain a 1% stock solution.

# **Bacterial Strain**

Clostridium chauvoei (jakari strain) isolated from clinically infected Zebu cattle was obtained in its lyophilised form from the National Veterinary Research Institute, Vom, Plateau state, Nigeria and used here.

### **Bacterial Cultivation**

Three media were used for cultivating Clostridium chauvoei (jakari strain) to isolate neuraminidase: reinforced clostridial medium (RCM), blood agar (BA) and cooked meat medium (CMM). Media preparation and bacterial cultivation were carried out as previously described. 10 All the procedures for microbiological asepsis were strictly maintained.

# Isolation of Neuraminidase from Culture

The growth medium CMM containing the cultivated bacteria was centrifuged at 9000 g for 40 min at 4°C. The supernatant (2L) containing crude neuraminidase was fractionated at 55-70% (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> saturation and dissolved in 20 mL of 50 mM acetate buffer pH 5.0. The enzyme solution was dialysed overnight against three changes of the same buffer. The dialysed enzyme was used for all the experiments. Total protein was quantified using Coomasie Brilliant Blue with bovine serum albumin as standard. 11

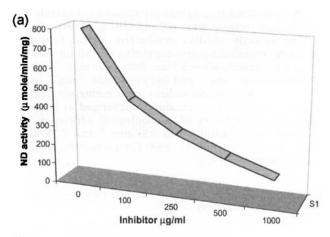
### **Enzyme Assay**

Neuraminidase was assayed using fetuin as substrate by quantifying the cleaved sialic acid. 12 Fetuin was purchased from Sigma Chemical Company, St. Louis, USA. Briefly, fetuin (98 mM) was dispensed into tubes (n = 100) and  $0.05 \,\mathrm{U/ml}$  of the purified culture supernatant containing Clostridium chauvoei (jakari strain) neuraminidase was added to it. Aliquots (0.25 ml) of sodium periodate was added to each sample and the mixture was shaken and incubated for 20 min in a water bath at 37°C after which 0.1 ml of sodium arsenite was added. The mixture was then shaken and placed in boiling water for 10 min when a pink colour appeared after boiling for 7.5–10 min.

The tubes containing all the mixtures above were cooled by placing them under a running tap. Thereafter, 2.5 ml of acid butanol was added to each tube and the mixture vigorously shaken. The tubes were then centrifuged at 1000 g for 5 min. The supernatant for each mixture was carefully aspirated into cuvettes using Pasteur pipettes and the absorbance was read against a blank using a Sp6-400 spectrophotometer at 549 nm. The amount of sialic acid cleaved from the substrate, was calculated using a sialic acid standard curve.

### Inhibition Studies

Various concentrations of the methanolic extracts  $100-1000 \,\mu g/ml$  were added to the reaction mixture containing 98 mM of fetuin, 0.05 U/ml of the Clostridium chauvoei neuraminidase and 50 mM acetate buffer pH 5.5 adjusted to 3 mL. The reaction was conducted at 37°C for 4h. After stopping the reaction, the released sialic acid was quantified as described previously and the results statistically analysed. 13 For each concentration of methanolic extract used, the test was repeated (n = 100) to check for reproducibility.



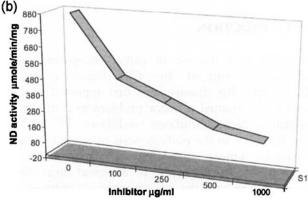


FIGURE 1 (a) Clostridium chauvoei Neuraminidase activity in the presence of different levels of the methanolic extract of Tamarindus indicus (b) Clostridium chauvoei Neuraminidase activity in the presence of different levels of the methanolic extract of Combretum fragrans.



The kinetic analysis of the enzyme was performed in the absence and presence of the extracts at pH 5.5 and 37°C. Fetuin at 9.8-49 mM was added to individual tubes and adjusted to 3 mL with acetate buffer, pH 5.5. Aliquots of the enzyme (0.05 U/mL) was added to the reaction mixture and the reaction was allowed to proceed for 2 h. On termination of the reaction, the released sialic acid was quantified as previously described.

# **RESULTS AND DISCUSSION**

The plants Tamarindus indicus and Combretum fragrans are used recurrently in the treatment of Blackleg.9 In the present work, a high level of inhibition of neuraminidase was observed by the methanolic extracts of Tamarindus indicus and Combretum fragrans. Mean neuraminidase activity was computed as enzyme activity ± Standard Deviation (SD) and there was inhibition of mean neuraminidase activity from  $6.1 \pm 4.2 \times 10^2$  to  $1.3 \times 10^2 \pm 1.6 \times 10^1$  and  $1.0 \times 10^2 \pm 4.9 \times 10^1$ µmole min<sup>-1</sup> in the presence of Tamarindus indicus and Combretum fragrans extracts respectively. Our results have shown at least a target of action by the plants which corroborates their use in traditional practice. Neuraminidases have been implicated in the pathologies of several diseases which include; cell invasion in Chagas disease,14 anaemia in trypanosomiasis<sup>15</sup> and viral invasion in Newcastle disease.16 The foregoing makes the sourcing of neuraminidase inhibitors mandatory for treatment and amelioration of clinical symptoms related to the physiological activity of the enzyme. In most cases the inhibitors are synthetic and indeed costly, e.g. Zanamivir used in the treatment of influenza virus infection.6 Moreover some of the synthetic inhibitors become less effective on account of mutations which are rampant in sialidases. This makes recourse to plants as source of neuraminidase inhibitors an appealing alternative, because their active compounds are synthesised in direct response to bacterial and viral invasion. In the present study, the two medicinal plants were tested because of their reported role in ameliorating blackleg infection in traditional veterinary practice. Both Tamarindus indicus and Combretum fragrans

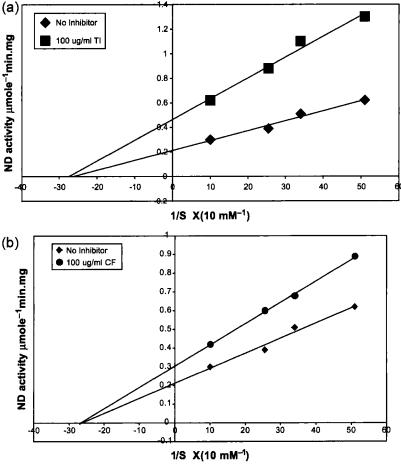


FIGURE 2 (a) Line weaver-Burk plots of Clostridium chauvoei neuraminidase catalysed hydrolysis of Fetuin in the absence and presence of 100 µg/ml of methanolic extract of Tamarindus indicus. (b) Line weaver-Burk plots of Clostridium chauvoei neuraminidase catalysed hydrolysis of Fetuin in the absence and presence of 100 µg/ml of methanolic extract of Tamarindus indicus.



342 N.M. USEH et al.

clearly showed a significant level of inhibition which was dose-dependent. As shown in Figure 1, the estimated IC<sub>50</sub> for Tamarindus indicus and Combretum fragrans extracts were 100 and 150 µg/ml respectively.

In order to determine neuraminidase inhibition by both plant extracts, a steady state analysis of neuraminidase activities was conducted at varied concentrations of fetuin, which assays the sialic acid cleaved by the enzyme. From the array of experiments, both Tamarindus indicus and Combretum fragrans exhibited non-competitive inhibition patterns against fetuin. As shown in the Figure 2a the extract from Tamarindus indicus reduced the V<sub>max</sub> from 500 \(\mu\)mole/min/mg to 240 \(\mu\)mole/min/mg. The  $V_{max}$  was reduced to 340  $\mu$ mole/min/mg when Combretum fragrans was used as inhibitor (Figure 2b). The K<sub>M</sub> value remained unchanged at 0.42 mM. Further analysis of the inhibition data showed a significant change in the index of physiological efficiency  $(V_{max}/K_M)$ . This was reduced from  $1.19\,\text{min}^{-1}$  to  $0.57\,\text{min}^{-1}$  and 0.75 min<sup>-1</sup> i.e. 50% and 40% inhibitions with Tamarindus indicus and Combretum fragrans as inhibitors respectively. It will be expedient to further characterize the active constituents of both plants which could be exploited in drug development.

# Acknowledgements

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## References

- Amstrong, H.L. and MacNamee, J.K. (1950) J. Am. Vet. Med. Assoc. 117, 212–214.
- [2] Osiyemi, T.I.O. (1975) Bull. Animal Health Prod. Africa 23(4), 367 - 370
- Muller, H.E. (1976) Zbl. Bakt. Hyg. 235, 106-110.
- [4] Esievo, K.A.N., Saror, D.I., Kolo, M.N. and Eduvie, L.O. (1986) 1. Comp. Pathol. 96, 95-96.
- [5] Von Itzstein, M., Wu, W.Y., Kok, G.B., Pegg, M.S., Dyasson, J.C. and Jin, B. (1993) Nature 363, 418-423
- [6] Hayden, F.G., Osterhalls, A.D., Treanor, J.J., Fleming, D.M., Aoki, F.V. and Nicholson, K.G. (1997) New Engl. J. Med. 337, 874-880.
- [7] Useh, N.M. (2002) "The production and characterization of neuraminidase from Clostridium chauvoei (Jakari strain)" M.Sc. Thesis, Ahmadu Bello University (Zaria, Nigeria), pp 224.
- [8] Jagun, A.G., Abdu, P.A., Mohammed, A.K., Alawa, C.B.I. and Omokanye, A.K. (1996) Survey of Ethnoveterinary Practices in Nigeria. Technical progress report on the survey conducted on ethnoveterinary practices in Nigeria, sponsored by International Development Research Centre (IDRC), Canada.
- [9] Abdu, P.A., Jagun, A.G., Gefu, G.O., Mohammed, A.K., Alawa, C.B.I. and Omokanye, A.T. (2000) A survey of ethnoveterinary practices of agropastoralists in Nigeria. In Ethnoveterinary Practices, Research and Development. Proceedings of an international workshop on ethnoveterinary practices held on 14-18th August, 2000, Kaduna, Nigeria. Edited by Jerome, G.O., Abdu, P.A. and Alawa, C.B.I. p163.
- [10] Dowell, V.R. and Hawkins, T.M. (1981) Laboratory Methods in Anaerobic Bacteriology, Centre for Disease Control (CDC) Laboratory manual (HHSPublication, Atlanta, Georgia), pp 1–96. [11] Scopes, K.R. (1984) Protein Purification Principles and Practices,
- Springer-Verlag; New York, pp 226.
- Aminoff, D. (1961) Biochemistry J. 81, 384-392.
- [13] Chatfield, C. (1983) Statistics for Technology. A Course in Applied Statistics, 3rded. (United Kingdom and Hall, London), 168-170, 186-190, pp 140-148. [14] Pereira-Chioccola, V.L. and Schenkman, S. (1999) *Biochem.*
- Soc. Trans. 27, 516-518.
- [15] Nok, A.J. and Balogun, E.O. (2003) J. Biochem. (Tokyo) 133, 725-730.
- [16] Oladele, S.B., Abdu, P.A., Nok, A.J., Esievo, K.A.N. and Useh, N.M. (2002) Veternarski arhiv. 72(4), 185-194.

